

## Synthesis of Arylalkylmonofluorophosphonates as Myo-Inositol monophosphatase Ligands.

## Laurent Schmitt, Nukhet Cavusoglu, Bernard Spiess, Gilbert Schlewer\*

Laboratoire de Pharmacochimie de la Communication Cellulaire, associé au CNRS Faculté de Pharmacie, 74, route du Rhin 67401 Illkirch, France

Received 12 February 1998; accepted 24 March 1998

Arylalkylmonofluorophosphonates Abstract: were prepared by condensation of lithium with the salt of diethyl 1-fluoro-1-(trimethylsilyl)methylphosphonate. Reduction and hydrolysis sequences gave the final products. These compounds do not inhibit the myo-inositol monophosphatase. © 1998 Published by Elsevier Science Ltd.

Myo-inositol monophosphatase seems to be a key enzyme in the cellular inositol phosphate cycle. Its uncompetitive inhibition by lithium salts, <sup>1-4</sup> modulating the cell signalling<sup>5</sup>, seems to be the mode of action of the manic depression therapy. These treatments are limited by the side effects which could lead to coma or death. <sup>6-9</sup> Discovery of new inhibitors of this enzyme could result in alternative therapies.

Structure-activity relationship studies were developed by differents groups. These studies relied on either systematic structural variations or took X-Rays, NMR and modelling analyses into account. Among numerous others, compounds 1-3 have demonstrated interesting binding potencies as competitive inhibitors.

These results suggested the synthesis of arylalkylmonofluorophosphonates analogues. Due to the fluor electronegativity, the monofluorophosphonate function has  $pK_A$  similar to the one of the corresponding phosphate. These analogues could be able to be complexed similarly to the natural ligands into the enzyme active site, but, could not be hydrolyzed. Moreover, the phenolic moiety could occupy the space area of the mechanistic water as for the phenolic group of 1.

Here, we report the synthesis of arylalkylmonofluorophosphonates 4.

$$\begin{array}{c} R_2 \\ R_3 \end{array} \begin{array}{c} R_1 \\ CH_2 \\ CH_2 \end{array} \begin{array}{c} O \\ CH_2 \\ CH_2 \end{array}$$

Compounds, where n = 0, were prepared starting from substituted benzaldehydes 5a-d (scheme 1). Reaction of these benzaldehydes with the lithium salt of diethyl 1-fluoro-1-(trimethylsilyl)-methylphosphonate <sup>19-23</sup> yielded a 1/1 cis/trans mixture of the alkenes 6a-d and 7a-d. Reduction of the double bond by hydrogenation using Pd/C as catalyst gave the corresponding fluoroalkanes 8a-d. The phosphonic esters were hydrolysed to give the free phosphonic acids 9a-d which were stabilized as ammonium salts. The phenolic group could be deprotected by treatment with BBr<sub>3</sub> 25 giving the phenol 10e-h, which were transformed to the free phosphonic acids 11e-h by means of trimethylsilyliodide. 24

 $Scheme~1: i)~ \text{LiTMSCFP(O)(OEt)}_2~1.2\text{eq, THF, -78°C,30 min} \rightarrow \text{RT}~30~\text{min, 73-81\%; j)}~ \text{H}_2,~ \text{Pd/C}\\ 10\%,~5~\text{atm, 15h, RT, 65-88\%; k)}~\text{TMSI, 4 eq, CH}_2\text{Cl}_2,~0^{\circ}\text{C, 30 min; l)}~ \text{BBr}_3,~ 3~\text{eq, CH}_2\text{Cl}_2,~-78^{\circ}\text{C, 2h} \rightarrow \text{RT}~12\text{h},~77-88\%; m)}~\text{TMSI, 4 eq, CH}_2\text{Cl}_2,~ 0^{\circ}\text{C, 30 min; PMBO} = p\text{-methoxybenzyloxy-.}^{26}$ 

For n = 2, the synthesis started from commarin 12 (scheme 2). Its reduction with NaBH<sub>4</sub> formed the 2-(3-hydroxy) propylphenol 13. The phenolic function was selectively protected as benzylether 14 and the primary alcohol was oxidized into aldehyde 15 by a Swern

reaction. <sup>27-29</sup> This aldehyde was submitted to the same fluorophosphonate coupling reaction as described above to give the alkenes 16 and 17. Hydrogenation of the double bond permitted the simultaneous deprotection of the phenol function yielding the fluorophosphonate 18.

Scheme 2: a) NaBH<sub>4</sub>, 2.5 eq, EtOH, 0°C 1h, RT 5h, reflux 5h 80%; b) NaH,1.2 eq, BnBr, 1.2 eq, DMF 0°C  $\rightarrow$  RT 2h 30 min, 91%; c) (COCl)<sub>2</sub>,3 eq, DMSO 4eq, Et<sub>3</sub>N, 12 eq, CH<sub>2</sub>Cl<sub>2</sub>,-78°C 45 min  $\rightarrow$  RT, 98%; d) LiTMSCFP(O)(OEt)<sub>2</sub>, 79%; e) H<sub>2</sub>, Pd/C, 98%; f) TMSI; g) BBr<sub>3</sub>; d  $\rightarrow$ g see scheme 1; Bn = benzyl.

Free phosphonic acid 19 was obtained by treatment of compound 18 with trimethylsilyliodide. Trimethylsilyliodide permitted to deprotect selectively the phosphonate esters of 16 and 17, and gave the derivative 20. But, the treatment of compound 16 and 17 by means of BBr<sub>3</sub> deprotected selectively the phenolic function and furnished compound 21 which, by treatment with trimethylsilyliodide gave the free phosphonic acid 22. The compounds 9a-d, 10e-h, 19, 20, and 22 were inactive toward the inhibition of the *myo*-inositol monophosphatase. A first molecular modelling approach showed that, even if the pK<sub>A</sub> could be similar to these of the natural ligands, the fluorine atom seemed less hydrophylic and unable to complex the magnesium cations and the amino-acid residue into the enzyme active site.

20

Acknowledgments. We thank Synthelabo Biomoleculaires and Dr A. Ganzhorn for providing us with human inositol monophosphatase and for assisting us for the enzymatic tests.

## References and notes

- 1. Hallcher, L.M.; Sherman, W.R. FEBS Lett. 1995, 361, 1-7.
- 2. Atack, J.R.; Broughton, H.B.; Pollack, S.J. FEBS Lett. 1995, 361, 1-7.
- 3. Leech, A.P.; Baker, G.R.; Shute, J.K.; Cohen, M.A.; Gani, D. Eur. J. Biochem. 1993, 212, 693-704.
- 4. Atack, J.R.; Broughton, H.B.; Pollack, S.J. Trends Neurosci. 1995, 18, 343-349.
- 5. Berridge, M.J.; Irvine, R.F. Nature, 1984, 312, 315-321.
- 6. Cade, J.F.J. Med. J. Aust. 1949, 2, 349-352.
- 7. Drummond, A.H. TIPS, 1987, 8, 129-133.
- 8. Berridge, M.J. J. Am. Med. Assoc. 1989, 262, 1834-1841.
- 9. Atack, J.R. Biol. Psychiatry, 1995, 37, 761-763.
- 10. Atack, J.R.; Fletcher, S.R. Drug of the Future, 1994, 19(9), 857-866.
- 11. Bone, R.; Springer, J.P.; Atack, J.R. Proc. Natl. Acad. Sci. USA, 1992, 89, 10031-10035.
- 12. York, J.D.; Ponder, J.W.; Chen, Z.W.; Mathews, F.S.; Majerus, P.W. *Biochemistry*, **1994**, 33, 13164-13171.
- 13. Bone, R.; Frank, L.; Springer, J.P.; Atack, J.R. Biochemistry, 1994, 33, 9468-9476.
- 14. Cole, A.G.; Gani, D. J. Chem. Soc., Perkin Trans. 1, 1995, 2685-2694.
- 15. Saudek, V.; Vincendon, P.; Do, Q.; Atkinson, R.A.; Sklenar, V.; Pelton, P.D.; Piriou, F.; Ganzhorn, A.J. Eur. J. Biochem. 1996, 240, 288-291.
- 16. Baker, R.; Carrick, C.; Leeson, P.D.; Lennon, I.C.; Liverton, N.J. J. Chem. Soc., Chem. Commun. 1991, 298-300.
- 17. MacLeod, A.M.; Baker, R.; Hudson, M. Med. Chem. Res. 1992, 2, 96-101.
- 18. Fletcher, S.R.; Baker, G.R.; Leeson, P.D.; Teall, M.; Harley, E.A.; Rogan, C.I. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 627-630.
- 19. Blackburn, G.M.; Taylor, G.E. J. Organomet. Chem. 1988, 348, 55-61.
- 20. Teulade, M.P.; Savignac, P. J. Organomet. Chem. 1988, 338, 295-303.
- 21. Patois, C.; Savignac, P. J. Chem. Soc., Chem. Commun. 1993, 1711-1712.
- 22. Nieschalk, J.; O'hagan, D. J. Chem. Soc., Chem. Commun. 1995, 719-720.
- 23. Waschbüsch, R.; Carran, J.; Savignac, P. Tetrahedron, 1996, 52, 14199-14216.
- 24. Groutas, W.C.; Felker, D. Synthesis, 1980, 861-868.
- 25. McOmie, J.F.W.; Watts, M.L.; West, D.E. Tetrahedron, 1968, 24, 2289-2296.
- 26. All the compounds gave satisfactory NMR (<sup>1</sup>H, <sup>31</sup>P, <sup>19</sup>F) analyses as well as elemental analyses.
- 27. Mancuso, A.J.; Huang, S.-L.; Swern, D. J. Org. Chem. 1978, 43, 2480-2482.
- 28. Corey, E.J.; Kim, C.U. J. Am. Chem. Soc. 1972, 94, 7586-7587.
- 29. Kulagowski, J.J. Tetrahedron Lett. 1989, 30, 3869-3872.